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Leading Edge

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XBP1 Links ER Stress to Intestinal Inflammation and Confers Genetic Risk for Human Inflammatory Bowel Disease p743

Arthur Kaser, Ann-Hwee Lee, Andre Franke, Jonathan N. Glickman, Sebastian Zeissig, Herbert Tilg, Edward E.S. Nieuwenhuis, Darren E. Higgins, Stefan Schreiber, Laurie H. Glimcher, and Richard S. Blumberg

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Inflammatory bowel disease (IBD) has been attributed to aberrant mucosal immunity to the intestinal microbiota. The transcription factor XBP1, a key component of the endoplasmic reticulum (ER) stress response, is required for development and maintenance of secretory cells and linked to JNK activation. We hypothesized that a stressful environmental milieu in a rapidly proliferating tissue might instigate a proinflammatory response. We report that Xbp1 deletion in intestinal epithelial cells (IECs) results in spontaneous enteritis and increased susceptibility to induced colitis secondary to both Paneth cell dysfunction and an epithelium that is overly reactive to inducers of IBD such as bacterial products (flagellin) and TNF α . An association of XBP1 variants with both forms of human IBD (Crohn's disease and ulcerative colitis) was identified and replicated (rs35873774; p value 1.6×10^{-5}) with novel, private hypomorphic variants identified as susceptibility factors. Hence, intestinal inflammation can originate solely from XBP1 abnormalities in IECs, thus linking cell-specific ER stress to the induction of organ-specific inflammation.

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 **Cover Caption**

On the cover: Coronins are highly conserved actin-binding proteins that localize to the leading edge of motile cells. In this issue, Cai et al. (pp. [828–842](#)) describe the mechanism of Coronin 1B function. This protein can target actin filament branches generated by the Arp2/3 complex, replace this complex, and produce a remodeled branch with altered geometry. This activity is antagonized by Cortactin, an actin-binding adaptor protein that protects the branches from disassembly. The illustration is an artistically rendered version of a dual-color kymograph used to demonstrate the temporal sequence of branch remodeling in the lamellipodia.



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