The Hsc70 co-chaperone CHIP targets immature CFTR for proteasomal degradation

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Protein quality control is a posttranslational process, and is essential for functional cell activity. Accumulation of misfolded polypeptides will cause many different kinds of diseases. There are two way to control the newly synthsized proteins' quality. One is refolding, the other is degradation. Here, this paper talks something about ubiquitination degradation.

### Abbreviation

CF	cystic-fibrosis, a most common genetic disease
CFTR	cystic-fibrosis transmembrane-conductance regulator
СНІР	C-terminal Hsp70 interacting protein (through a set of tetratricorepeat motifs)
U-box	ubiquitin ligase domain
TPR	tetratricorepeat / tetratricopeptide repeats
ALLN	N-acetyl-leucyl-norleucinal
Hsc70	a cytosol Heat Shock Chaperone (?)

First, I want to list some abbreviations I will use during this presentation.

CFTR is a plasma membrane ion channel that exposes the majority of its mass in the cytosol. We can see its distribution from the microscopy picture.

U-box, is a 75 amino acid domain first identified in UFD2, which is a yeast E4 enzyme. Recent study showed that U-box containing proteins functions as an E3 enzyme, and takes part in the ubiquitination and degradation.

I googled, but not sure what is Hsc70, or are there any different between Hsp70 and Hsc70. I thought there are the same.



<u>Co-immunoprecipitation</u> is an extension of the immunoprecipitation technique which is used to determine whether a given protein interacts physically with another given protein. Cell extracts containing the presumed interaction complex are first immunoprecipitated with antibody against one of the proteins. The material identified by this means is then tested for the presence of the other protein by immunoblotting with a specific antibody.

A typical ABC transporter consists of four domains: two highly hydrophobic domains, each with six putative membrane-spanning segments that somehow form the translocation pathway, and two ATP-binding catalytic domains (or cassettes). In some cases the two halves of the transporter are formed by a single polypeptide (*as shown*), whereas in other cases they are formed by two separate polypeptides.

Co-chaperone, I thought it is a chaperone of another chaperone.



- There are hundreds of different mutations that can result in CF with varying degrees of severity.
- The most common mutation, ΔF508, results in a single amino acid deletion. CFTR-ΔF508 is thought to fold much more slowly than the wildtype CFTR protein.

Cystic fibrosis is the most common disease in U.S. And it relates to the mutation of CFTR.

One common mutation, delta F508 is a single amino acid deletion. And the author use this mutation as a compare in this paper, and I thought we can rule out the possible mechanism of how this mutation causes disease.

As in other literature, they showed that CFTR-delta F508 folds much slower than wild type, combine with the conclusion from this paper, I thought that the aberrant folding of CFTR-delta F508 leads to the aberrant degradation, which may interfere with CFTR's function.



As there are so many co-IP in this paper, I would like to introduce the main structure of this paper first.

First, use immunofluorescent microscopy, they found CFTR/CHIP/Hsc70 are co-localized.

Second, they found that this degradation is not interfered by other chaperons, and have a high specificity to target immaturely glycosylated CFTR, which is called B form.

Third, the authors found that the degradation of immature CFTR is through ubqiuitinating, probably in proteasome.

Finally, they mentioned that the U box of CHIP is essential for the degradation, while TPR motif is necessary but not sufficient.

### CHIP co-localizes with CFTR and Hsc70 at the ER



+, means treated with ALLN, an inhibitor of the 20s proteasome. If the green and red co-localize, it will show yellow.

Elevated CHIP levels, through the expression of exogenous Myc–CHIP, prevent the cell-surface localization of CFTR and cause green fluorescent protein (GFP)–CFTR14 to accumulate in a perinuclear location.

When cells are treated with the proteasome inhibitor ALLN, degradation of misfolded CFTR is inhibited and it accumulates in intracellular inclusions termed aggresomes. In ALLN-treated cells, the localization of CHIP is altered, and it can be colocalized in the aggresome with proteasomes, Hsc70 and CFTR.

Thus, CHIP has the potential to interact with CFTR in the ER or in aggresomes. It is important to note, however, that CFTR degradation initiates in the ER and that CFTR is not normally detected in aggresomes. Thus, CHIP probably functions at the ER to influence GFP–CFTR localization.

### Elevation of CHIP reduces CFTR-C, but not accumulates CFTR-B



### Degradation CFTR may involve the proteasome



#### CFTR promotes the degradation of newly synthesized CFTR



#### TPR & U-box are necessary not sufficient for CHIP's function



#### CFTR's degradation by CHIP is not interfered with ER insertion



## CHIP does not influence the degradation of apolipoprotein B48



### Hsc70/CHIP recognizes CFTR conformation-specifically



### CHIP does not interfere the binding of Hsc70 to CFTR



# CHIP & Hsc70 function independently to CFTR



### CHIP forms complex with Hsc70 and subunit from the proteasome



SDS,解聚各类的蛋白作用。

# The U-box is required for CHIP to promote CFTR ubiquitination



### Overexpression of CHIP U-box blocks CFTR degradation





### Conclusions

- CHIP functions with Hsc70 to target the immature CFTR for degradation by ubiquitination.
- The U-box appears essential for this process.
- Hsc70/CHIP may function specifically in ER quality control.

? How distinguish the protein for degradation

In this paper, they found out that:

I thought that because CFTR delta-F508 has a lower folding velocity, which will accumulated unfolded protein, and may intrigue the degradation.

But there are still many questions remained. One big question is that how CHIP/Hsc70 complex distinguish between misfolded protein and canonical protein.



Thanks for your attention.